

---

# Folium Menthae Piperitae

## Definition

Folium Menthae Piperitae consists of the dried leaves of *Mentha* × *piperita* L. (Lamiaceae) (1–3).

## Synonyms

*Mentha piperita* (L.) Huds., *M. piperita* Stokes, *M. balsamea* Willd. (1, 4).

## Selected vernacular names

Amentha, american mint, balm mint, brandy mint, cabra-caa, curled mint, doun menta piperita, hierbabuena, hortela pimenta, Katzenkraut, lamb mint, la menta, lamint, menta piemonte, mentea peperina, mentha pepe, menthe, menthe anglaise, menthe poivrée, moto yuyo, nána, ni naa, ni'na el fulfully, pepermin, pepper mint, peppermint, Pfefferminze, Pfefferminzblätter, piperita, pudeena, pum hub, yerba mota (1, 4, 5).

## Geographical distribution

Commercially cultivated in eastern and northern Europe and the United States of America, and is found in Africa (1, 3, 6, 7).

## Description

A perennial herb, 30–90 cm high. Stems square erect or ascending, branched, the upper portion always quadrangular. Leaves opposite, petiolate, ovate-oblong to oblong-lanceolate, serrate, pointed; dark green on the upper surface. Flowers purplish, occur in thick, terminal, spicoid racemes of verticillasters; each flower shows a tubular calyx with 5 sharp, hairy teeth, a purplish, irregular, 4-cleft corolla, 4 short stamens, a 4-celled ovary and a projecting style ending in a bifid stigma. Fruit consists of 4 ellipsoidal nutlets (1, 7, 8).

## Plant material of interest: dried leaves

### General appearance

Green to greenish-brown. Leaves whole, broken or cut; thin, fragile; whole leaf 3–9 cm long and 1–3 cm wide, often crumpled. Lamina oval or lanceolate; apex acuminate; margin sharply dentate; base asymmetrical. Venation pinnate,

prominent on the lower surface, with lateral veins leaving the midrib at an angle of about 45°. Lower surface slightly pubescent and secretory trichomes visible under a hand lens as bright yellowish points. Petiole grooved, usually up to 1 mm in diameter and up to 1 cm long (2).

### **Organoleptic properties**

Odour: characteristic, penetrating; taste: characteristic, aromatic (2).

### **Microscopic characteristics**

Upper epidermis composed of large, clear epidermal cells with sinuous, vertical walls and possessing few or no stomata, few glandular trichomes present; palisade parenchyma, comprising a layer of columnar cells rich in chloroplasts; spongy parenchyma, of 4–6 layers of irregularly shaped chloroplastid-containing cells and intercellular air-spaces. Lower epidermis of small epidermal cells with sinuous, vertical walls and numerous diacytic stomata; in the region of veins and midrib, exhibits non-glandular and glandular trichomes as outgrowths; non-glandular trichomes uniseriate, papillose, 1–8-celled; glandular trichomes have 1–2-celled stalk and 1–8-celled glandular head containing the essential oil. Calcium oxalate crystals absent; pollen grains spheroidal and smooth (1, 4, 7, 8).

### **Powdered plant material**

Brownish-green. Fragments of leaf tissue with cells of epidermis having sinuous walls, cuticle striated over the veins, diacytic stomata present predominantly on the lower epidermis; epidermis fragments from near leaf margin with isodiametric cells showing distinct beading and pitting in anticlinal walls; covering trichomes short, conical, unicellular, bicellular or elongated, uniseriate multicellular (3–8 cells) with striated cuticle. Glandular trichomes of 2 types: either with unicellular base with small, rounded, unicellular head 15–25 µm in diameter; or with unicellular base with enlarged, oval multicellular head 55–70 µm in diameter composed of 8 radiating cells; dorsoventral mesophyll fragments with a single palisade layer and 4–6 layers of spongy parenchyma; yellowish crystals of menthol under the cuticle of secretory cells. Calcium oxalate crystals absent (1, 2).

### **General identity tests**

Macroscopic and microscopic examinations, and thin-layer chromatography (1, 2). Gas chromatography of the steam-distilled essential oil (9).

### **Purity tests**

#### **Microbiological**

Tests for specific microorganisms and microbial contamination limits are as described in the WHO guidelines on quality control methods for medicinal plants (10).

### **Foreign organic matter**

Not more than 5% stems, the diameter of which must be not more than 1.5 mm; not more than 8% leaves showing brown stains due to *Puccinia menthae* (2); not more than 2% other foreign matter (2).

### **Total ash**

Not more than 15% according to the *European pharmacopoeia* (2); not more than 12% according to the *African pharmacopoeia* (4).

### **Acid-insoluble ash**

Not more than 1.5% (2).

### **Pesticide residues**

The recommended maximum limit of aldrin and dieldrin is not more than 0.05 mg/kg (2). For other pesticides, see the *European pharmacopoeia* (2), and the WHO guidelines on quality control methods for medicinal plants (10) and pesticide residues (11).

### **Heavy metals**

For maximum limits and analysis of heavy metals, consult the WHO guidelines on quality control methods for medicinal plants (10).

### **Radioactive residues**

Where applicable, consult the WHO guidelines on quality control methods for medicinal plants (10) for the analysis of radioactive isotopes.

### **Other purity tests**

Sulfated ash, water-soluble extractive, alcohol-soluble extractive, and loss on drying tests to be established in accordance with national requirements.

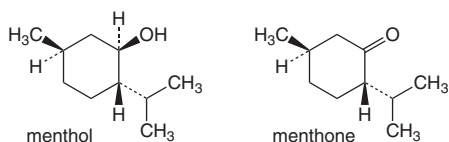
## **Chemical assays**

Whole and cut leaves contain not less than 1.2% and 0.9% (v/w) essential oil, respectively, determined as described in the *European pharmacopoeia* (2).

## **Major chemical constituents**

The major constituent of the leaves is the essential oil (0.5–4%), which contains menthol (30–55%) and menthone (14–32%). Menthol occurs mostly in the free alcohol form, with small quantities as the acetate (3–5%) and valerate esters. Other monoterpenes present include isomenthone (2–10%), 1,8-cineole (6–14%),  $\alpha$ -pinene (1.0–1.5%),  $\beta$ -pinene (1–2%), limonene (1–5%), neomenthol (2.5–3.5%) and menthofuran (1–9%) (2, 4, 6, 12, 13).

The structures of the major monoterpenes, menthol and menthone, are presented below.



## Medicinal uses

### *Uses supported by clinical data*

None.

### *Uses described in pharmacopoeias and in traditional systems of medicine*

Symptomatic treatment of dyspepsia, flatulence and intestinal colic (1, 3, 14, 15).

### *Uses described in folk medicine, not supported by experimental or clinical data*

As an emmenagogue, vermifuge, lactation enhancer and sedative. Also used to treat bronchitis, bacillary dysentery, diabetes, diarrhoea, dysmenorrhoea, fevers, hypertension, jaundice, nausea, pain, and respiratory and urinary tract infections (5).

## Pharmacology

### *Experimental pharmacology*

#### **Antimicrobial activity**

Extracts of Folium Menthae Piperitae have antibacterial and antiviral activity in vitro. Addition of ground leaves to the agar medium inhibited the growth of *Salmonella typhimurium*, *Staphylococcus aureus* and *Vibrio parahaemolyticus* at concentrations of 0.1–2.0% (w/v) (16). Aqueous and ethanol extracts of the leaves reduced the number of plaques of the rinderpest virus at concentrations of 4–8 mg/ml (17). Aqueous extracts of the leaves demonstrated activity against the following viruses in egg and cell culture: Newcastle disease, herpes simplex, vaccinia, Semliki Forest and West Nile (18).

#### **Smooth muscle contraction**

A 31% ethanol extract of the leaves inhibited both acetylcholine- and histamine-induced smooth muscle contractions in guinea-pig ileum in vitro at a concentration of 10 ml/l (19, 20). The results were similar to those obtained with 0.13 mg atropine (19). An aqueous flavonoid fraction isolated from a leaf

extract inhibited barium chloride-induced muscle contractions of guinea-pig ileum in vitro at a concentration corresponding to 0.5 g leaves/ml (21).

### **Choleretic activity**

Injection of a leaf infusion (0.5 ml) or a flavonoid fraction (equivalent to 3.3 g leaves/kg body weight) increased the amount of bile acids in cannulated rats and dogs (dose 0.4 mg/kg body weight) (21, 22). A mixture of flavonoids, isolated from the leaves, had choleretic activity in dogs (2 mg/kg body weight) (23). Flavomentin, a flavonoid isolated from the leaves, stimulated bile secretion and the synthesis of bile acids in dogs (2 mg/kg body weight) (24). Intra-gastric administration of a 30% ethanol extract of the leaves to rats (1 ml/kg body weight) increased bile flow by 43%. The extract did not induce sedation in mice at doses up to 10 ml/kg body weight (20).

### **Anti-oedema activity**

Topical application of a methanol leaf extract to mice (2.0 mg/ear) inhibited ear oedema induced by 12-*O*-tetradecanoylphorbol-13-acetate (25).

### **Analgesic activity**

Intra-gastric administration of a 30% ethanol extract of the leaves inhibited phenylbenzoquinone-induced writhing in mice (ED<sub>50</sub> 2.1 ml/kg body weight) (20).

### **Toxicology**

Intra-gastric administration of a leaf extract (50 g leaves infused with 500 ml hot water for 10 minutes, then spray-dried) to 12 mice (4 g/kg body weight as a single dose) did not result in central nervous system depression, toxic effects or mortality (26).

### ***Clinical pharmacology***

None.

### **Contraindications**

No information available.

### **Warnings**

No information available.

### **Precautions**

#### ***General***

Patients with gallstones should not use *Folium Menthae Piperitae* unless under medical supervision (15).

### **Other precautions**

No information available on precautions concerning drug interactions; drug and laboratory test interactions; carcinogenesis, mutagenesis, impairment of fertility; teratogenic and non-teratogenic effects in pregnancy; nursing mothers; or paediatric use. Therefore, Folium Menthae Piperitae should not be administered during pregnancy or lactation or to children without medical supervision.

### **Adverse reactions**

No information available.

### **Dosage forms**

Dried leaves (2, 3). Tincture and infusions (6). Store in a well-closed container, protected from light (2).

### **Posology**

(Unless otherwise indicated)

Daily dosage: 1–3 g crude drug three times daily (14, 27). Infusion: pour 150 ml hot water over 1.5–3.0 g (one tablespoon) dried leaves, steep for 10 minutes, strain and drink three times daily between meals (6, 15, 28). Tincture: 2–3 ml (1:5, 45% ethanol) three times daily (14).

### **References**

1. *African pharmacopoeia*. Vol. 1, 1st ed. Lagos, Organization of African Unity, Scientific Technical & Research Commission, 1985.
2. *European pharmacopoeia*, 3rd ed. Strasbourg, Council of Europe, 1996.
3. *British herbal pharmacopoeia*. London, British Herbal Medicine Association, 1996.
4. Blaschek W et al., eds. *Hagers Handbuch der pharmazeutischen Praxis. Folgeband 2: Drogen A–K*, 5th ed. Berlin, Springer-Verlag, 1998.
5. Farnsworth NR, ed. *NAPRALERT database*. Chicago, University of Illinois at Chicago, IL, February 9, 1998 production (an online database available directly through the University of Illinois at Chicago or through the Scientific and Technical Network [STN] of Chemical Abstracts Services).
6. Bisset NG. *Herbal drugs and phytopharmaceuticals*. Boca Raton, FL, CRC Press, 1994.
7. Youngken HW. *Textbook of pharmacognosy*, 6th ed. Philadelphia, PA, Blakiston, 1950.
8. Evans WC. *Pharmacognosy*, 14th ed. London, WB Saunders Co., 1996.
9. *Pharmacopoeia Hungarica*, 7th ed. Budapest, Hungarian Pharmacopoeia Commission, Medicina Konyvkiado, 1986.
10. *Quality control methods for medicinal plant materials*. Geneva, World Health Organization, 1998.
11. *Guidelines for predicting dietary intake of pesticide residues*, 2nd rev. ed. Geneva, World Health Organization, 1997 (document WHO/FSF/FOS/97.7).
12. Bruneton J. *Pharmacognosy, phytochemistry, medicinal plants*. Paris, Lavoisier, 1995.
13. Samuelsson G. *Drugs of natural origin, a textbook of pharmacognosy*. Stockholm, Swedish Pharmaceutical Press, 1992.

14. Bradley PR, ed. *British herbal compendium. Vol. 1.* Bournemouth, British Herbal Medicine Association, 1992.
15. Blumenthal M et al., eds. *The complete German Commission E monographs.* Austin, TX, American Botanical Council, 1998.
16. Aktug SE, Karapinar M. Sensitivity of some common food-poisoning bacteria to thyme, mint and bay leaves. *International Journal of Food Microbiology*, 1986, 3:349–354.
17. Alwan AH et al. Antiviral activity of some Iraqi indigenous plants. *International Journal of Crude Drug Research*, 1988, 2:107–111.
18. Herrmann EC Jr, Kucera LS. Antiviral substances in plants of the mint family (Labiatae). III. Peppermint (*Mentha piperita*) and other mint plants. *Proceedings of the Society for Experimental Biology and Medicine*, 1967:874–878.
19. Forster HB et al. Antispasmodic effects of some medicinal plants. *Planta Medica*, 1980, 40:309–319.
20. Leslie GB. A pharmacometric evaluation of nine Bio-Strath herbal remedies. *Medita*, 1978, 8:3–19.
21. Lallement-Guilbert N, Bézanger-Beauquesne L. Recherches sur les flavonoides quelques Labiées médicinales (romarin, menthe poivrée, suage officinale). *Plantes médicinales et Phytothérapie*, 1970, 4:92–107.
22. Steinmetzer K. Experimentelle Untersuchungen über Cholagoga. *Wiener Klinische Wochenschrift*, 1926, 39:1418–1422, 1455–1457.
23. Pasechnik IK. Study of choleric properties specific to flavonoids from *Mentha piperita* leaves. *Farmakologija Toksikologija*, 1966, 21:735–737.
24. Pasechnik IK, Gella EV. Choleric preparation from peppermint. *Farmatsevtichnyi Zhurnal (Kiev)*, 1966, 21:49–53.
25. Yasukawa K et al. Inhibitory effect of edible plant extracts on 12-O-tetradecanoylphorbol-13-acetate-induced ear edema in mice. *Phytotherapy Research*, 1993, 7:185–189.
26. Della Loggia R et al. Evaluation of some pharmacological activities of a peppermint extract. *Fitoterapia*, 1990, 61:215–221.
27. Wichtl M. Pfefferminzblätter. In: Wichtl M, ed. *Teedrogen*, 2nd ed. Stuttgart, Wissenschaftliche Verlagsgesellschaft, 1989:372–374.
28. *ESCOP monographs on the medicinal uses of plant drugs.* Fascicule 3. Devon, European Scientific Cooperative on Phytotherapy, 1997.